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## RESEARCH ARTICLE

# Host range of antibiotic resistance genes in wastewater treatment plant influent and effluent

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**One sentence summary:** epicPCR was utilized to link antibiotic resistance genes to bacterial host species in two wastewater purification plants.

**Editor:** Tong Zhang

## ABSTRACT

Wastewater treatment plants (WWTPs) collect wastewater from various sources for a multi-step treatment process. By mixing a large variety of bacteria and promoting their proximity, WWTPs constitute potential hotspots for the emergence of antibiotic resistant bacteria. Concerns have been expressed regarding the potential of WWTPs to spread antibiotic resistance genes (ARGs) from environmental reservoirs to human pathogens. We utilized epicPCR (Emulsion, Paired Isolation and Concatenation PCR) to detect the bacterial hosts of ARGs in two WWTPs. We identified the host distribution of four resistance-associated genes (*tetM*, *int1*, *qacEΔ1* and *bla<sub>OXA-58</sub>*) in influent and effluent. The bacterial hosts of these resistance genes varied between the WWTP influent and effluent, with a generally decreasing host range in the effluent. Through 16S rRNA gene sequencing, it was determined that the resistance gene carrying bacteria include both abundant and rare taxa. Our results suggest that the studied WWTPs mostly succeed in decreasing the host range of the resistance genes during the treatment process. Still, there were instances where effluent contained resistance genes in bacterial groups not carrying these genes in the influent. By permitting exhaustive profiling of resistance-associated gene hosts in WWTP bacterial communities, the application of epicPCR provides a new level of precision to our resistance gene risk estimates.

**Keywords:** epicPCR; antibiotic resistance; wastewater treatment; horizontal gene transfer

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## INTRODUCTION

The emergence and spread of antibiotic resistance genes (ARGs) among pathogenic and non-pathogenic bacteria has been a rising threat in recent decades. Among suspected sites for resistance transmission are wastewater treatment plants (WWTPs) where wastewater from various sources, including municipalities, hospitals and industries, is mixed and treated in a multi-step purification process. As the microbes from different sources are in close contact during the purification process, WWTPs have been suggested to be hotspots for the emergence and dissemination of antibiotic-resistant bacteria (Rizzo et al. 2013a). The wastewater treatment process creates conditions that may favor horizontal gene transfer with high bacterial densities, stress caused by pollutants such as heavy metals and antibiotics, and biofilms formed during the purification process (Karkman et al. 2017).

Currently, studies evaluating the scope and frequency of antibiotic resistance in WWTPs are largely based on the use of quantitative PCR (qPCR) (Auerbach, Seyfried and McMahon 2007; Karkman et al. 2016; Karkman et al. 2017), metagenomics (Bengtsson-Palme et al. 2016) or cultivation. With qPCR, ARGs can be quantified in the studied environment, and recent advances in the use of high-throughput qPCR arrays (Stedtfeld et al. 2008) have enabled the detection and quantification of hundreds of genes simultaneously (Karkman et al. 2016; Muziasari et al. 2016; Muurinen et al. 2017). Nevertheless, the information about gene-carrying species is missing, and whether the ARG carriers are potential pathogens or harmless environmental bacteria remains unresolved. Cultivation-based approaches, although widely used in clinical microbiology, are limited as only a small fraction of microbes can be grown on laboratory media (Amann, Ludwig and Schleifer 1995). In recent years, the use of metagenomics in ARG research has increased (Yang et al. 2014; Bengtsson-Palme et al. 2016), but the approach still has several challenges. The relatively low abundance of resistance genes and their frequent presence on mobile genetic elements, which can be distributed in several different species, as well as difficulties in the assembly of mosaic genetic elements, complicate linking taxonomic information to specific ARGs. The application of Inverse-PCR (Pärnänen et al. 2016) has enabled circumventing problems in the assembly of ARGs and their genetic context. However, genes that are localized on mobile genetic elements are still difficult to assign to a specific species due to their distribution in more than one taxa.

Differences in the taxonomy of ARG carrying bacteria in WWTP influent and effluent could indicate potential transfer events during the wastewater purification process. To study this, we utilized epicPCR (Spencer et al. 2016) to gain a comprehensive understanding of the species carrying ARGs in the raw influent and purified effluent of WWTPs. epicPCR has been utilized so far for determining the bacterial host diversity of the dissimilatory sulfite reductase gene, *dsrB* (Spencer et al. 2016). epicPCR requires a priori knowledge of the sequences of the target genes of interest, provided for this study by prior qPCR experiments (Karkman et al. 2016). epicPCR workflow begins by the encapsulation of a sufficient cell number ( $2 \times 10^7$ ) individually into hydrogel beads, after which a gene of interest (here, ARGs) is linked to the 16S rRNA gene of the host bacterium with the use of concatenating PCR. These concatenated amplicons are subsequently sequenced using high-throughput sequencing technologies, primarily Illumina MiSeq. Finally, the hosts of the ARG are identified bioinformatically based on 16S rRNA gene sequence.

For the selection of genes to be used in epicPCR, we utilized data gathered with qPCR (Laht et al. 2014) and qPCR array (Karkman et al. 2016) from the Viikinkäki WWTP over four seasons. The following genes abundant in the WWTPs were selected for the study: *tetM* encodes a protein protecting from inhibition of translation by tetracycline at the ribosome (Roberts 2005). Tetracycline resistance genes are common in feces (Hu et al. 2013) and thus found from WWTPs. *bla<sub>OXA-58</sub>* is a beta-lactamase gene conferring resistance to carbapenems (Poirel et al. 2005) by hydrolyzing the compound, and it is clinically significant as carbapenems belong to last-resort antibiotics. It has been detected in WWTPs (Caucci et al. 2016; Karkman et al. 2016), and our interest was the host range of this ARG. *int1* was chosen as antimicrobial resistance genes may be captured and thus spread via class I integrons that recruit genes into a series of gene cassettes ensuring their expression and are frequently present in mobile elements (Gillings et al. 2008). Class 1 integrons have a broad host range and have been detected in both Gram-positive and Gram-negative bacteria including a diverse array of obligatory and opportunistic human and zoonotic pathogens (Nandi et al. 2004; Gillings et al. 2015). Lastly, *qac $\Delta$ 1*, a biocide resistance gene, is linked to clinical class I integrons containing various ARGs (Gillings et al. 2008).

Here, our aim was to determine the bacterial hosts of the ARG of interest and find differences in the host distribution of the ARGs in raw inflow and effluent water at two municipal wastewater treatment plants.

## MATERIALS AND METHODS

### Sample collection

Samples were collected from two different municipal treatment plants in Helsinki and Espoo, Finland. Viikinkäki is the largest WWTP in Nordic countries processing wastewater from 800 000 residents and from the region's industry (annually 100 million m<sup>3</sup> of wastewater). Approximately 15% of the wastewater is from industry and 85% is domestic wastewater. The Suomenoja WWTP processes the wastewaters of 310 000 residents (annually 35 million m<sup>3</sup> of wastewater) and around 8% of the wastewater originates from industry. 24 h collection samples were taken on three subsequent days from influent and from effluent water in September 2016. The collection ended in the morning, and the samples were transported to the laboratory within 2 h.

Cells for epicPCR were collected from 2 ml of the influent wastewater by centrifuging for 5 min at  $11\,000 \times g$  and from 200 ml of effluent wastewater by centrifuging for 40 min at  $8000 \times g$ . After discarding the supernatant, the remaining cell pellets were resuspended in 20% glycerol, flash frozen with liquid nitrogen and stored in  $-80^\circ\text{C}$ .

For DNA extractions, 50 ml of influent wastewater and 200 ml of effluent water were filtered through 0.22  $\mu\text{m}$  polycarbonate filters (diameter 47 mm, Whatman) in triplicate. The filters for influent water were cut in half with a sterile scalpel and forceps, and filters were transferred to Mobio PowerWater DNA Isolation Kit (Mobio Laboratories, CA, USA) bead beating tubes and stored in  $-20^\circ\text{C}$ .

### DNA extraction and 16S rRNA gene PCR

DNA from the filters was extracted with Mobio PowerWater DNA Isolation Kit following manufacturer's instructions. The DNA was quantified with Qubit Broad-Range Assay Kit

(Thermo Fisher Scientific, Waltham, MA, USA). PCR amplification of the V3–V4 region of the 16S rRNA gene was performed in two steps. The first round of amplification was done with the primers 341F1-4 and 785R1-4 that contain partial Illumina TruSeq adapter sequences in the 5' ends (Table 1) using Phusion polymerase with GC buffer and 2.5% DMSO (New England Biolab, MA, USA). The cycling conditions were as follows: initial denaturation at 98°C, followed by 15 cycles at 98°C for 10 s, 65°C for 30 s and 72°C for 10 s, and a final extension for 5 min at 72°C. The amount of template DNA used was approximately 25 ng. The PCR run included a PCR negative control without template DNA. The PCR products were purified with exonuclease I (Thermo Fisher Scientific, Waltham, MA, USA) and thermosensitive alkaline phosphatase (FastAP; Thermo Fisher Scientific, Waltham, MA, USA). A second PCR round was performed with full-length TruSeq P5 and Index containing P7 adapters. The cycling conditions were identical to the previous ones except that 18 instead of 15 cycles were run. The final PCR products were purified with Agencourt® AMPure® XP magnetic beads (Beckman Coulter, CA, USA), pooled and sequenced on the Illumina MiSeq platform at the Institute of Biotechnology, University of Helsinki, Finland. All 16S rRNA gene sequences have been deposited in the European Nucleotide Archive (accession no. PRJEB23695).

### Primer design

ARGs were selected based on previous results (Karkman et al. 2016) and targeted using previously published primers (Table 1). Because of the melting temperatures of the targeted ARGs, the pH' primer (Table 1) was used as the reverse primer for amplifying the 16S rRNA gene, differing from the protocol used by Spencer et al. (2016), who used a primer with a lower melting temperature. Also, the primer 785R from the original protocol used in nested PCR was modified (Table 1), and short TruSeq adapter sequences were added to the nested PCR primers.

### epicPCR

The droplets for epicPCR were done for cells stored in glycerol as described by Spencer et al. (2016) with the following modifications: for bead formation, 100 µl autoclaved MilliQ water (Millipore), 100 µl 30% BIS/acrylamide (29:1, Bio Rad, CA, USA) and 25 µl 10% APS (Sigma, MO, USA) were used. A 35 µm cell strainer (Falcon, NY, USA) was used in bead straining. The prevalence of beads containing several cells and the distribution of beads containing a single bacterium versus empty beads were estimated by staining cells with SybrGreenII (Thermo Fisher Scientific, Waltham, MA, USA) and microscopied under a compound microscope (Zeiss Axioskop 2 plus, Oberkochen, Germany).

The first PCR was done as described in Spencer et al. (2016) with the following exceptions: Phusion Hot Start Flex DNA Polymerase (NEB) was used with the GC buffer and 55°C annealing temperature and without water, BSA or Tween-20 as in Spencer et al. (2016). After bead purification, nested PCR was conducted in quadruplicate. For some genes, a semi-nested approach was used with primers including TruSeq adapters (Table 1). In nested PCR, the annealing temperature was set to 55°C. PCR reactions were purified with the Agencourt AMPure XP purification system (Beckman Coulter, Inc., CA, USA) followed by eight cycles of PCR with TruSeq P5 and Index primers containing P7 adapters to prepare the samples for sequencing. The samples were sequenced on the Illumina MiSeq platform at the Institute of Biotechnology, University of Helsinki, Finland.

All epicPCR sequences have been deposited in the European Nucleotide Archive (accession no. PRJEB23695).

### 16S rRNA gene data analysis

For 16S rRNA reads were joined with Pear (Zhang et al. 2014) with default options and quality trimmed using USEARCH -fastq\_filter command with -fastq\_maxee 1 and -fastq\_minlen 350 parameters. Unique sequences were identified with the UPARSE pipeline (Edgar 2013) with -derep\_fulllength command. OTUs were clustered, chimeras removed and reads were mapped to reference sequences with the -cluster\_otus command with -minsize 2 parameter and -usearch\_global command with -id 0.97 parameter. Taxonomic classification of OTUs was done using the classify\_seqs command in mothur (Schloss et al. 2009) using the RDP naïve Bayesian Classifier (Wang et al. 2007) against the Silva 128 database (Quast et al. 2013) with classifier cutoff = 60.

The community composition between the two plants and the two different stages were compared with permutational multivariate analysis of variance using the function Adonis from the vegan package (Oksanen et al. 2017) in R (R Core team 2017). Shannon index and inverse Simpson index describing the alpha diversity of the microbial communities were calculated with phyloseq (McMurdie and Holmes 2013) as well as beta diversity with the Bray–Curtis dissimilarity index.

### epicPCR data analysis

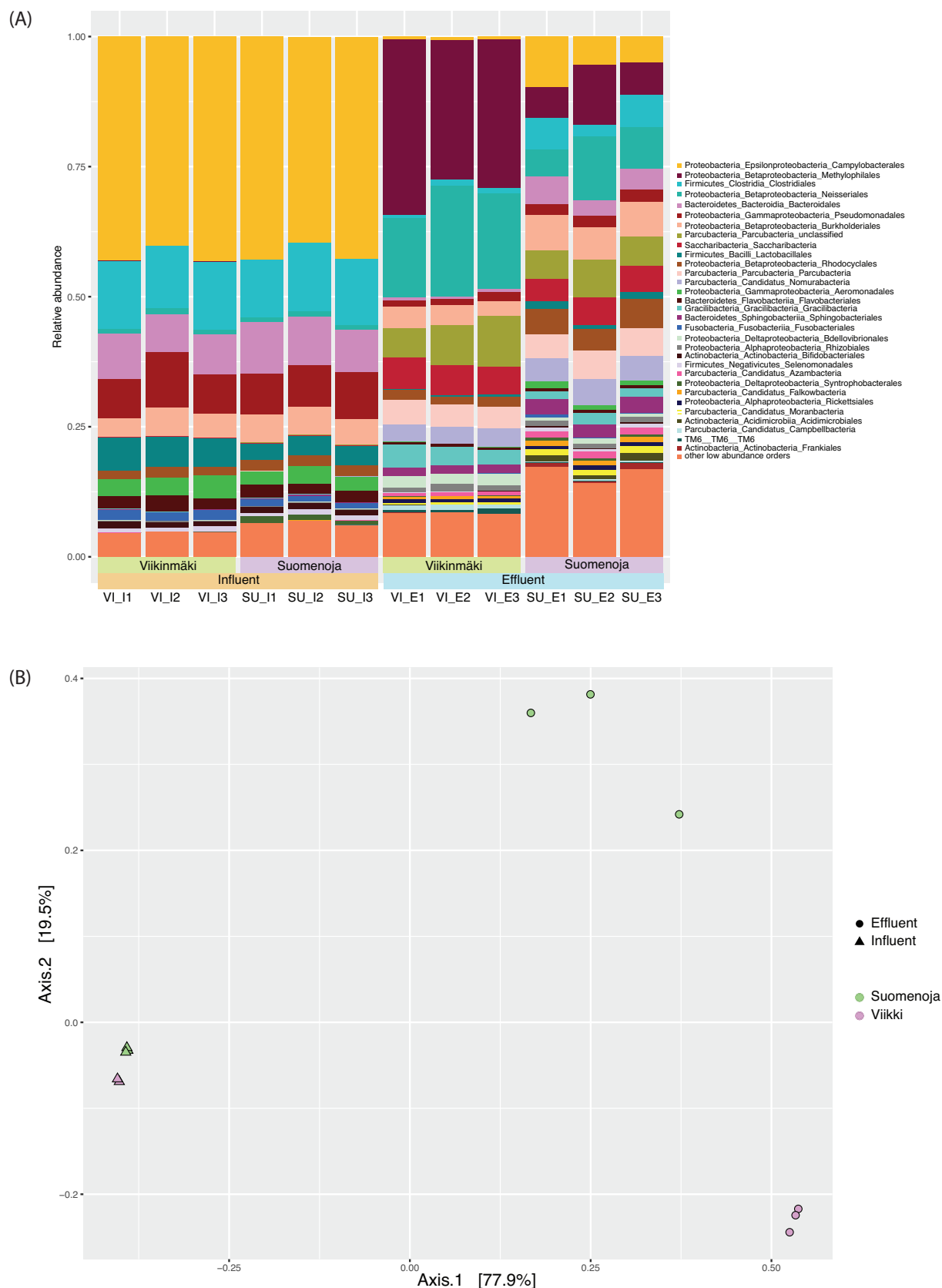
The paired end reads were joined with Pear (Zhang et al. 2014) and binned based on the ARG of interest. Short reads (<400 bp) were removed with cutadapt (Martin 2011) and the good quality reads were split into 16S rRNA and ARG sequences with Prinseq (Schmieder and Edwards 2011) based on the functional gene sequence. 16S rRNA gene OTUs (97%) were identified using UPARSE with the -cluster\_otus command with -minsize 2 parameter and -usearch\_global command with -id 0.97 parameter. The OTUs were assigned taxonomies in mothur (Schloss et al. 2009) using the RDP naïve Bayesian classifier (Wang et al. 2007) against Silva 128 database (Quast et al. 2013) with classifier cutoff = 60. ARGs were also clustered to OTUs but using 100% identity and annotated with BlastN (Altschul et al. 1990) to verify that the correct gene was amplified. Results for a gene in a sample were taken into consideration when there were >57 reads after quality filtering. The average number of reads was 17 888.

For 16S rRNA gene OTUs, a representative sequence from each bacterial family was aligned with PyNAST (Caporaso et al. 2010), and a phylogenetic tree of the representative sequences was built with Fasttree (Price, Dehal and Arkin 2009). The presence/absence of an ARG in the bacterial family of origin of OTUs was visualized in iTol (Letunic and Bork 2016). A gene was considered present in the OTU of interest when found in all samples passing the quality filtering step (2/2 or 3/3) from the influent/effluent in a plant of interest.

## RESULTS

### Microbial community composition in the WWTPs

The bacterial community composition in the Viikimäki and Suomenoja WWTPs was characterized by similar influent composition. In contrast, the effluent communities of the two plants differed substantially (PERMANOVA with Bray–Curtis dissimilarity: plant  $P = 0.007$ , phase  $P = 0.0001$ , plant: phase  $P = 0.003$ ; Fig. 1a and b). The main species in the influent belonged to the



**Figure 1.** (A) Order level classification of the 16S rRNA gene sequences. Microbial community composition was analyzed from both plants (VI = Viikimäki, SU = Suomenoja) in the influent and effluent water on three subsequent days. (B) Principal coordinate analysis (PCoA) plot from the OTU relative abundance with the Bray–Curtis dissimilarity. The influent clusters from two plants clustered together, whereas the effluent samples were more distinct at the studied plants.



**Table 1.** Primers used in the study.

Gene name	sequence 5'-3' <sup>a) b)</sup>	Target	Ref.
341F1	ATCTACACTCTTTCCCTACACGACGCTCTTCCGATCTCCTACGG GNGGCWGCAG	16S rRNA gene	Herlemann et al. (2011)
341F2	ATCTACACTCTTTCCCTACACGACGCTCTTCCGATCTgtCCTACGG GNGGCWGCAG	16S rRNA gene	Herlemann et al. (2011)
341F3	ATCTACACTCTTTCCCTACACGACGCTCTTCCGATCTagagCCTAC GGGNGGCWGCAG	16S rRNA gene	Herlemann et al. (2011)
341F4	ATCTACACTCTTTCCCTACACGACGCTCTTCCGATCTtagtgCCT ACGGNGGCWGCAG	16S rRNA gene	Herlemann et al. (2011)
785R1	GTGACTGGAGTTCAGACGTGTGCTCTTCCGATCTGACTACHVGG GTATCTAATCC	16S rRNA gene	Herlemann et al. (2011)
785R2	GTGACTGGAGTTCAGACGTGTGCTCTTCCGATCTaGACTACHVG GGTATCTAATCC	16S rRNA gene	Herlemann et al. (2011)
785R3	GTGACTGGAGTTCAGACGTGTGCTCTTCCGATCTtctGACTACHVG GGTATCTAATCC	16S rRNA gene	Herlemann et al. (2011)
785R4	GTGACTGGAGTTCAGACGTGTGCTCTTCCGATCTctgagtGACTA CHVGGGTATCTAATCC	16S rRNA gene	Herlemann et al. (2011)
tetM.F1	CATCATAGACACGCCAGGACA	tetM epic forward	Karkman et al. (2016)
tetM.R1.F2'	GWATTACCGCGGCKGCTGCTGTTTGGATTACAATTTCCGC	tetM epic linker	Tamminen et al. (2011)
TetM.F3.TS	ATCTACACTCTTTCCCTACACGACGCTCTTCCGATCTGCAATTCT ACTGATTTCTGC	tetM epic nested	Tamminen et al. (2011)
qacEΔ1.F1	TCGCAACATCCGCATTAAAA	qacEΔ1 epic forward	Eckert, Gautier and Arlet (2006)
qacEΔ1.R1.F2'	GWATTACCGCGGCKGCTGATGGATTTCAGAACCAGAGAAAGA AA	qacEΔ1 epic linker	Karkman et al. (2016)
qacEΔ1.F1.TS	ATCTACACTCTTTCCCTACACGACGCTCTTCCGATCTTCGCAA CATCCGCATTAAAA	qacEΔ1 epic nested	Karkman et al. (2016)
int1-a.F1	CGAAGTCGAGGCATTTCTGTGTC	int1 epic forward	Muziasari et al. (2014)
int1-a.R1.F2'	GWATTACCGCGGCKGCTGGCCTTCCAGAAAACCGAGGA	int1 epic linker	Muziasari et al. (2014)
int1-a.F1.TS	ATCTACACTCTTTCCCTACACGACGCTCTTCCGATCTCGAAGT CGAGGCATTTCTGTGTC	int1 epic nested	Muziasari et al. (2014)
bla <sub>OXA-58</sub> .F1	ACAGGCACTGTAGATGCTTG	bla <sub>OXA-58</sub> epic forward	Karkman et al. (2016)
bla <sub>OXA-58</sub> .R1.F2'	GWATTACCGCGGCKGCTGTGTGCTGAGCATAGTATGAG	bla <sub>OXA-58</sub> epic linker	Karkman et al. (2016)
bla <sub>OXA-58</sub> .F3.TS	ATCTACACTCTTTCCCTACACGACGCTCTTCCGATCTTCGGTCTA AATGCGTGCCAT	bla <sub>OXA-58</sub> epic nested	Karkman et al. (2016)
pH'	AAGGAGGTGATCCAGCCGCA	16S rRNA gene	Edwards et al. (1989)

<sup>a</sup>Lower case nucleotides in primers 341F and 785R are introduced for mixing in sequencing.

<sup>b</sup>Nucleotides in bold mark the 16S rRNA gene sequence (in R1.F2') or the short Illumina TrueSeq adapter (in nested primers).

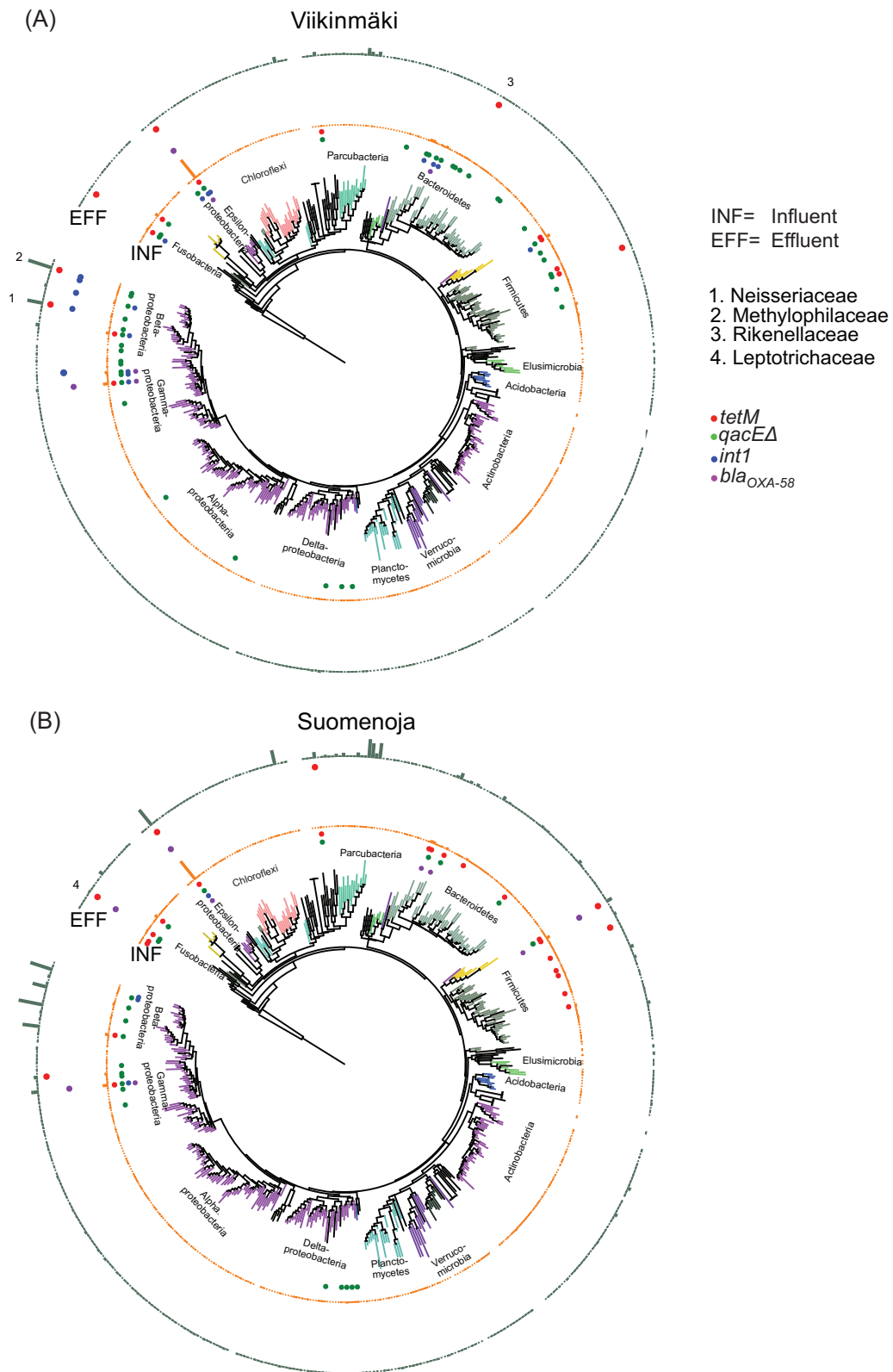
bacterial order Campylobacteriales with 39%–43% of the reads assigned to this order. Other orders with high relative abundance in the influent included Clostridia (12.55 ± 1.6%), Bacteroidales (8.53 ± 1.0%) and Pseudomonadales (8.43 ± 1.2%). However, the community profiles differed in the WWTP effluents (Fig. 1a): in Viikinmäki, the main orders were Methylophilales (29.70 ± 3.6%), Neisseriales (18.31 ± 3.0%) and unclassified Paracubacteria (7.72 ± 2.0%); in Suomenoja, there were no clear dominant species but the diversity was higher (Fig. S1, Supporting Information) and the orders with the highest relative abundance were Neisseriales (8.48 ± 3.5%), Methylophilales (7.8 ± 3.2%) and Campylobacteriales (6.68 ± 2.65%).

### Resistance-associated gene hosts as determined by epicPCR

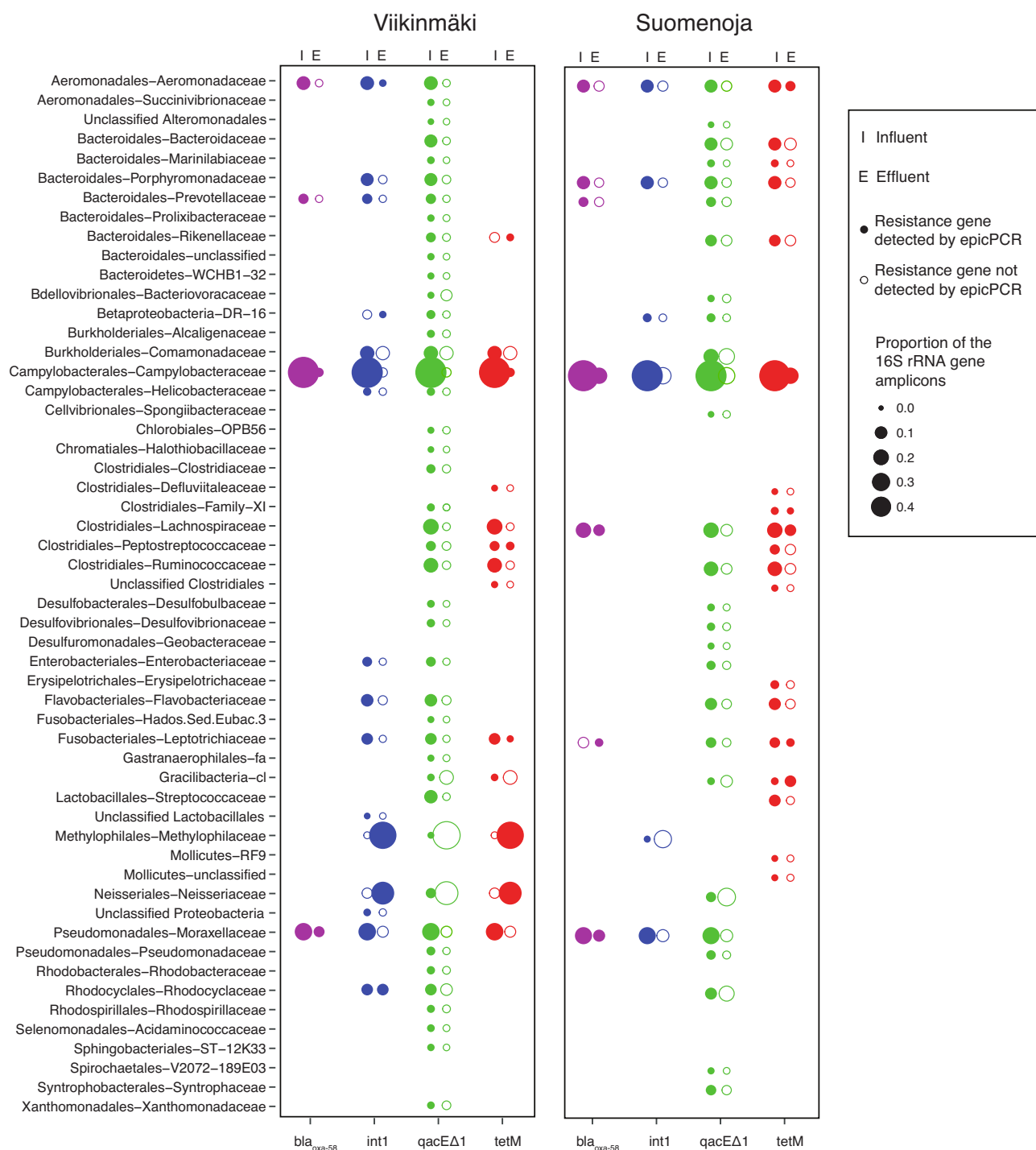
Two ARGs, the tetracycline resistance gene, *tetM*, and beta-lactamase gene, *bla<sub>OXA-58</sub>*, together with the class I integron integrase *intI* and biocide resistance gene, *qacEΔ1* (Table 1), were analyzed with epicPCR from the two WWTPs. We modified the epicPCR protocol from the published one (Spencer et al. 2016) by using 16S rRNA gene primers with a higher melting temperature.

All of the studied genes were detected in both influent and effluent. However, the OTUs carrying ARGs and ARG associated genes differed from the most abundant OTUs in the total community. There were also differences in the bacterial genera carrying ARGs between the two WWTPs (Figs 2 and 3; Tables S1 and S2, Supporting Information). Due to the inherent qualitative nature of epicPCR resulting from a large number of cycles in two PCR steps, we analyzed the epicPCR results using presence/absence data as opposed to a quantitative approach. When comparing the number of OTUs associated with the targeted ARGs and ARG associated genes detected in the epicPCR analyses to the total number of OTUs detected in the 16S rRNA sequence library, we found that the ratio (Table 2) of these two was generally higher in the raw influent wastewater than in the more diverse (Fig. S1, Supporting Information) effluent water.

In Viikinmäki, *tetM* was found in 13 bacterial genera in the influent (Table S1, Supporting Information), whereas it was observed in six genera in the effluent. A similar profile was observed in Suomenoja, although the number of *tetM* carrying genera was higher in Suomenoja than in Viikinmäki with 28 genera in the influent and nine in the effluent wastewater (Table S1, Supporting Information). These genera were distributed in families of the phyla Bacteroidetes, Firmicutes, Proteobacteria,



**Figure 2.** Presence of ARGs in microbial families in the two wastewater purification plants. The phylogenetic tree was constructed from family-level OTUs based on 16S rRNA gene amplicon sequences. An ARG was considered to be present in a bacterial genus when found in 2/2 or 3/3 samples (depending on whether all three samples passed the QC step) from the influent or effluent water at the same plant. The inner circle (INF) denotes the influent and the outer circle (EFF) the effluent water. The results are presented at family level and bars in the INF and EFF rings display relative abundance.



**Figure 3.** Bacterial families with resistance gene detected by epicPCR in influent and/or effluent of the two plants (Viikinmäki and Suomenoja). Relative abundance of the family in total 16S rRNA gene amplicon sequencing is shown with the size of the sphere. Open sphere: gene not detected with epicPCR, closed sphere: gene detected with epicPCR.

Tenericutes, Fusobacteria and Gracilibacteria (Fig. 2). The host range of *tetM* was broader than that of *bla*<sub>OXA-58</sub>. In Viikinmäki five and in Suomenoja seven genera were observed to carry the gene in the influent, and two and eight, respectively, in the effluent. The phyla containing the genera carrying *bla*<sub>OXA-58</sub> included members of Bacteroidetes, Firmicutes, Fusobacteria and Proteobacteria (Fig. 2). The gene *qacEΔ1* had the broadest host range observed with epicPCR: in the influent *qacEΔ1* was found in 84

and 44 genera in Viikinmäki and Suomenoja, respectively, and in the effluent in four genera in both treatment plants (Table S1, Supporting Information). For the *int1* gene in the Viikinmäki WWTP, there was larger diversity in the influent than in the effluent, with 17 genera associated with the gene in the influent and only six in the effluent. In Suomenoja, there were seven genera carrying *int1* in the influent but we were not able to detect the gene in the effluent water with epicPCR.



**Table 2.** Number of OTUs detected in epicPCR versus 16S rRNA gene amplicon sequencing.

sample	tetM	bla <sub>OXA-58</sub>	int1	qacEΔ1
VI.I1	1.14%	0.69%	0.55%	6.57%
VI.I2	1.04%	0.52%	1.51%	–
VI.I3	2.95%	0.73%	1.77%	7.04%
VI.O1	0.68%	0.22%	0.00%	0.19% <sup>a</sup>
VI.O2	–	0.24%	0.34%	0.64%
VI.O3	0.32%	–	0.45%	–
Sample	tetM	bla <sub>OXA-58</sub>	int1	qacEΔ1
SU.I1	2.90%	0.40%	–	2.78%
SU.I2	–	0.36%	0.32%	–
SU.I3	3.89%	2.22%	0.59%	5.31%
SU.O1	0.65%	–	–	–
SU.O2	–	0.23%	–	0.36% <sup>a</sup>
SU.O3	1.62%	0.29%	–	–

<sup>a</sup>When only one of the three replicates was successful, the data is not presented in Figs 2 and 3.

## DISCUSSION

EpicPCR was originally used to profile the host distribution of the dissimilatory sulfite reductase gene, *dsrB*, in linker PCR with the 16S rRNA gene in the protocol by Spencer *et al.* (2016). Here, for the first time, we applied epicPCR to link ARGs and class I integron associated genes with the taxonomic information of the bacteria carrying them. We were able to detect differences in ARG and integron carrying bacterial genera in the wastewater influent and treated effluent at the two purification plants studied. We also analyzed the total bacterial communities in the studied samples and found that both the total community and the observed ARG and integron carrying genera changed during the wastewater treatment process.

We were able to observe putative HGT events using epicPCR. In the Suomenoja effluent water, *bla*<sub>OXA-58</sub> gene was found to be associated with a Leptotrichiaceae OTU belonging to Fusobacteria, while the gene was not detected in this phylum in the influent. Similarly, *tetM* was observed in the effluent in an uncultured member of Rikenellaceae with low relative abundance in the total community and two abundant members of Methylophilaceae and Neisseriaceae families, while *tetM* was not detected in these families in the raw influent (Figs 2 and 3; Table S2, Supporting Information). Due to the highly variable OTU abundances in the samples, we cannot definitively conclude whether the differences in the host species distribution were due to horizontal gene transfer or the detection limit of the method. Clearly, the abundance of reads attained in 16S rRNA gene sequencing does not limit the detection of hosts in epicPCR (Fig. 2) as bacterial families with both low and high relative abundance were linked to the four studied genes. However, mechanistic proof of gene transfer during the treatment process would require further methodological advancements in *in situ* HGT studies.

The total relative abundance of ARGs has been shown to decline during the wastewater purification process (Yang *et al.* 2014; Bengtsson-Palme *et al.* 2016; Karkman *et al.* 2016) but the relative abundance of some genes is enriched in the effluent bacterial community (Mao *et al.* 2015; Munck *et al.* 2015). The conditions in the purification process are often considered to favor horizontal gene transfer (Rizzo *et al.* 2013b). However, actual data supporting active HGT in WWTPs is rather scarce. It remains unknown whether the species carrying ARGs are the

same before and after the treatment process but are at low abundance and thus below detection limit, or whether the wastewater treatment process favors horizontal gene transfer of ARGs. The latter is observed when the taxa that carry ARGs are different in the effluent from those in the influent. However, reliable association of ARG carriage with horizontal gene transfer events requires that all the cases of ARG carriage are detected. This warrants further investigation into factors influencing the detection limit of epicPCR.

The gene *tetM* was carried in partly different bacterial families in the influent and effluent wastewater. Members from the families Methylophilaceae, Neisseriaceae and Rikenellaceae harbored *tetM* in effluent water, although they were not observed to carry the gene in the influent suggesting possible HGT events. Of these families, Neisseriaceae is known to contain pathogenic species (Rotman and Seifert 2014), whereas Rikenellaceae and Methylophilaceae are typical bacteria in the wastewater purification process. The *tetM* gene is commonly localized on a conjugative transposon, Tn916, with a broad host range (Roberts and Mullany 2009) possibly explaining the observed diversity of the *tetM* carrying taxa in effluents.

The gene *bla*<sub>OXA-58</sub>, a plasmid-borne gene providing resistance to carbapenem, is commonly detected in *Acinetobacter baumannii*. *A. baumannii* is an opportunistic pathogen responsible for a range of nosocomial infections, especially in intensive care units (Heritier *et al.* 2005). Here, the gene was linked to *Acinetobacter*, but also to gammaproteobacterial genera including *Tolomonas*, *Aeromonas* and *Enhydrobacter*. In addition, *bla*<sub>OXA-58</sub> was linked to *Arcobacter*, the family Lachnospiraceae belonging to Clostridia, the family Leptotrichiaceae belonging to Fusobacteria and several Bacteroidales families. These groups contain bacteria found in feces and include pathogenic species as well as environmental bacteria. Although the *bla*<sub>OXA-58</sub> gene has been considered to be affiliated with *A. baumannii* and a few other *Acinetobacter* species (Coelho *et al.* 2006), it was recently characterized in the enterobacterium *Proteus mirabilis* isolated from a plasmid in clinical samples (Lange *et al.* 2017). In *P. mirabilis*, the gene was found in a plasmid with a different genetic context than in *A. baumannii*. epicPCR linked the gene to several genera, suggesting the possibility of this gene having been transferred to more taxa than originally thought. However, the number of species the gene was linked to decreased during the wastewater treatment process. Despite the decrease in genera, we observed *bla*<sub>OXA-58</sub> to be associated with the family Leptotrichiaceae in the

effluent in Suomenoja WWTP, although the gene was not associated to the family in the influent.

The four studied genes had differences in their host range, but interestingly, *Arcobacter* was found to carry all of the investigated genes in the effluent water. *Arcobacter* belongs to Campylobacteria and species of the genus are considered to be emergent pathogens (Miller et al. 2007). The species in this genus have been found to carry several ARGs in their genome, and in our study, the genus harbored the widest range of genes. Recently, a member of this genus common in WWTPs has been found to carry several ARGs (Millar and Raghavan 2017). However, there are only 14 genome sequences of *Arcobacter* in the NCBI genome database (2017–11-22) and their diversity in the environment is largely unknown. The results suggest that *Arcobacter* species might be important carriers of ARGs in WWTP environments.

This study demonstrates that epicPCR can be used to determine the host range of ARGs in complex environmental samples. Adding more primers to the used set is straightforward, enabling the analysis of the host ranges of multiple ARGs in complex communities at single cell level in a high-throughput manner.

## SUPPLEMENTARY DATA

Supplementary data are available at [FEMSEC](#) online.

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